

# Repurposing cannabidiol (CBD) as novel antimicrobial candidate targeting methicillin-resistant and vancomycin-intermediate *Staphylococcus aureus*

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## INTRODUCTION

Infections caused by methicillin-resistant *Staphylococcus aureus* (MRSA) are responsible for high morbidity and mortality worldwide. The horizontal acquisition of vancomycin-resistance genes results in the rise of vancomycin-intermediate *S. aureus* (VISA). Herein in this study, we report the antimicrobial and anti-biofilm activity of cannabidiol (CBD) and CBD formulations containing minor cannabinoids against multidrug-resistant (MDR) MRSA and VISA strains harboring genetically defined resistance mechanisms.

## METHODS

### Isolation of non-psychoactive cannabinoids

The non-psychoactive cannabinoids were extracted from *Cannabis sativa* plants using commercial extraction methods. The CBD was further purified using distillation, extraction and crystallization techniques. The minor cannabinoids were further purified from crude extract using chromatographic techniques.



### Characterization of cannabinoid compositions

The chemical composition of cannabinoids was quantified by using high performance liquid chromatography (HPLC) and UV spectrometry.

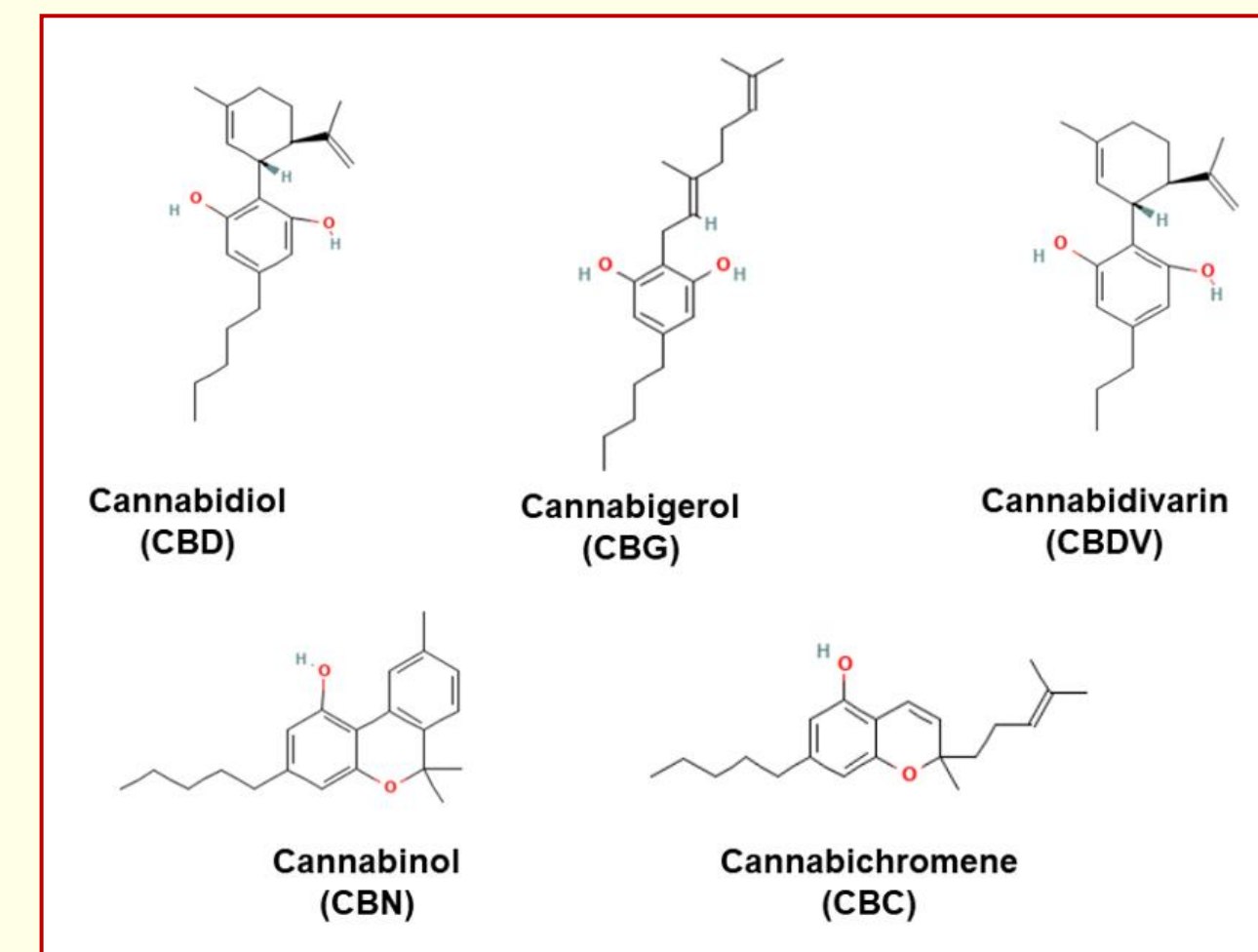
### Minimal inhibitory concentration determination

The minimal inhibitory concentration (MIC; µg/ml) of pure CBD isolate as well as CBD formulations containing variable amounts of minor cannabinoids (CBC, CBN, CBDV and CBG) was determined using the broth microdilution method using *S. aureus* strains with genetically defined resistance mechanisms.

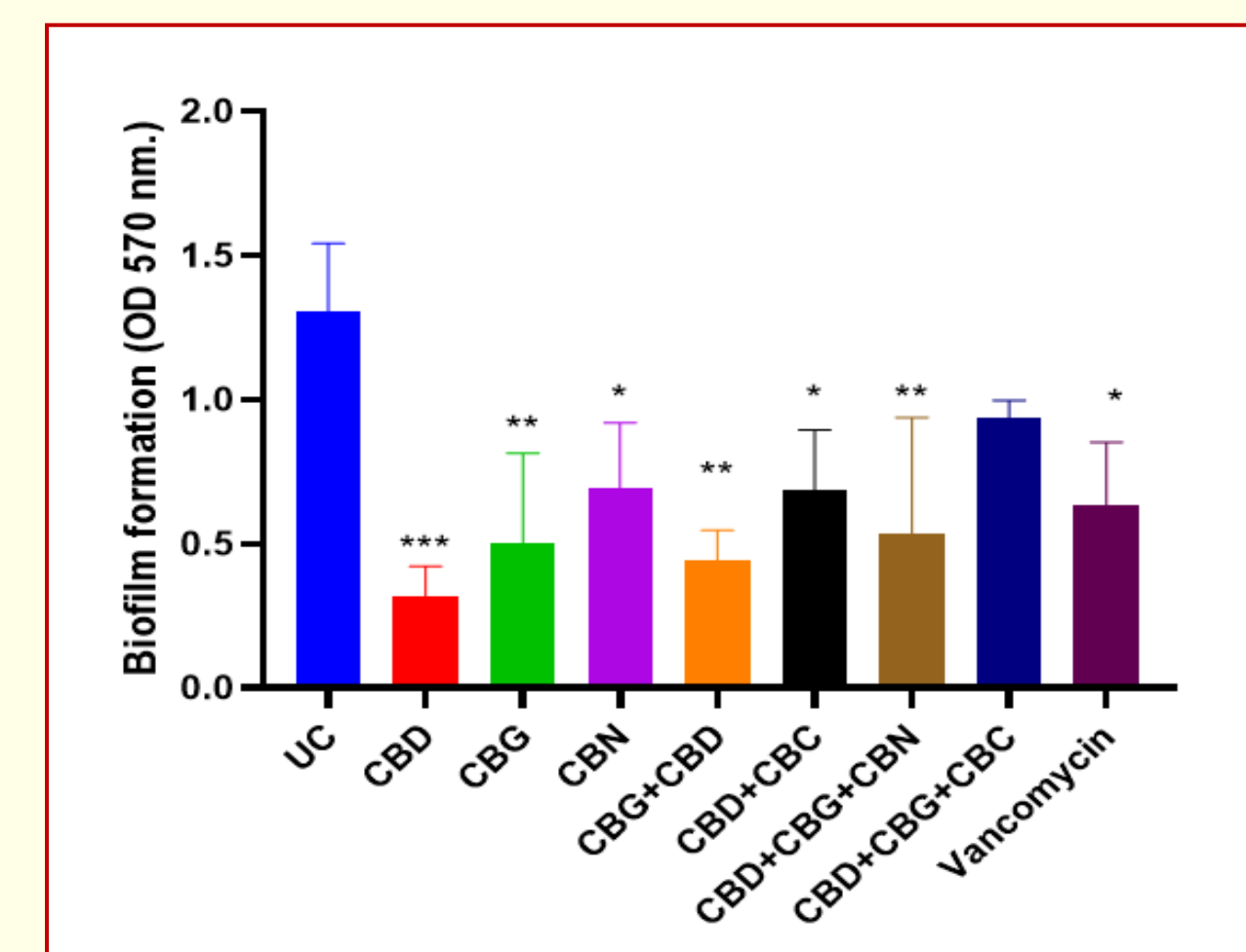
### Biofilm formation assay

The effect of CBD alone and in combination with minor cannabinoids on *S. aureus* biofilms formation and development was characterized by using spectrophotometric crystal violet (CV) assay.

## RESULTS



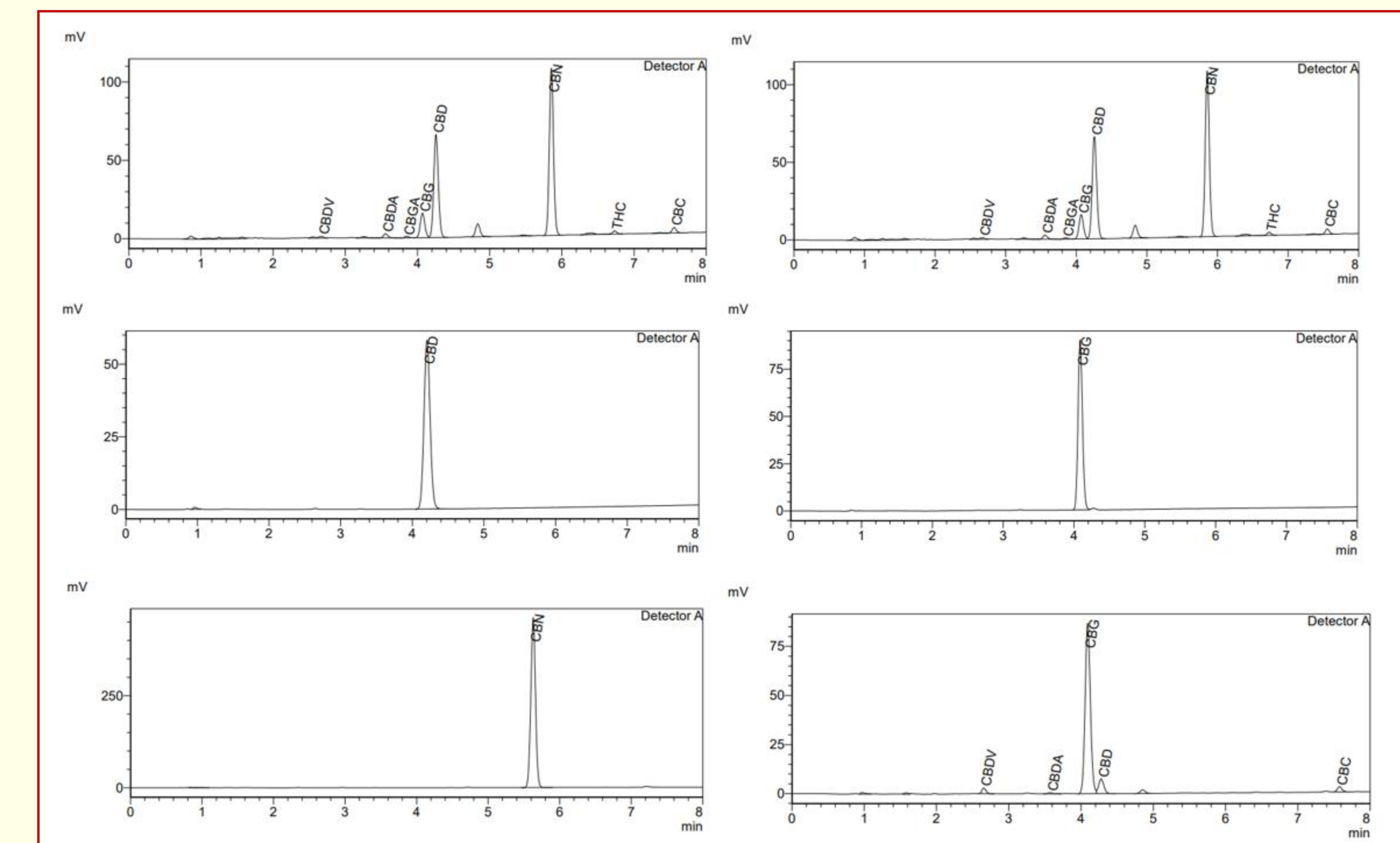
**Figure 1.** The chemical structures of major, non-psychoactive cannabinoids isolated from *Cannabis sativa* plant.



**Figure 2.** The effect of cannabinoids on *S. aureus* MRSA biofilm formation. The biofilms were exposed with 1X MIC concentration of test compounds for 24 hours.

**Table 1.** The minimal inhibitory concentration of non-psychoactive cannabinoids against *Staphylococcus aureus* isolates with genetically defined resistance mechanisms.

Pathogen	Resistance mechanisms	MIC							
		CBD	CBG	CBN	CBG+CBD	CBD+CBC	CBD+CBG+CBN	CBD+CBG+CBC	Vancomycin
<i>S. aureus</i> Pan-S	<i>blaZ</i>	1	1	2	8	32	64	8	0.5
<i>S. aureus</i> MRSA	<i>mecA</i>	2	4	2	16	16	64	8	2
<i>S. aureus</i> VISA	<i>aac(6)-aph(2'')</i> , <i>aadD</i> , <i>erm(A)</i> , <i>mecA</i> , <i>spc</i> , <i>tet(M)</i>	1	4	2	32	32	128	2	8



**Figure 3.** The detection and characterization of major cannabinoids extracted from *C. sativa*. The cannabinoids of interest were further purified by using liquid chromatography and their chemical purity was confirmed by HPLC.

Pure CBD, CBG and CBN demonstrated satisfactory and selective antimicrobial activity (2-4 µg/ml) against all tested MRSA and VISA strains and was comparable or greater than vancomycin (1-2 µg/ml and 4 µg/ml, respectively).

## CONCLUSIONS

These results demonstrated the promising translational application of CBD as a novel candidate for future development of novel antimicrobial candidates targeting multidrug-resistant *Staphylococcus aureus* and their biofilms.

## REFERENCES

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